

## Review

# A Critical Mini-Review on Atmospheric Ozone Mediated Alterations of Bioaerosols and Their Effects on Human Health

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**Abstract:** Bioaerosols are airborne particles that contain microorganisms and their derivatives, attracting much attention recently due to global epidemic of COVID-19. In fact, characteristics of bioaerosols can be significantly influenced by pollutants in air. As one of the most common ambient air pollutants, ozone (O<sub>3</sub>) may influence the characteristics of bioaerosols and finally affects their health effects. However, the interaction association between the atmospheric ozone pollution and bioaerosols are poorly understood. In this critical mini-review, recent research about the influences of O<sub>3</sub> on biological components, physical characteristics, bio-activity, evolution of community structure as well as health risk of bioaerosols is reviewed. In addition, this mini-review also highlights that atmospheric O<sub>3</sub> may play a potential role to boost the spread of antibiotics resistance genes to some extent, which warns the public to properly control atmospheric O<sub>3</sub> and bioaerosol pollutions synchronously.

**Keywords:** bioaerosol; ozone pollution; alteration of characteristics; change of community structure; health risk

## 1. Introduction

Bioaerosols are airborne particles that contain microorganisms and their derivatives, such as bacteria, fungi, viruses, endotoxin, mycotoxins, pollen and so on [1,2]. Bioaerosols exist outdoor and indoor with the bacterial and fungi concentration ranging from tens to thousands and several to hundreds CFU·m<sup>-3</sup>, respectively, depending on seasons and locations [3,4]. The size distribution of bioaerosols shows large span from 10 nm to 100 μm, which size of airborne bacteria majorly distributes within 1.0–7.0 μm, while fungi within 1.0–11.0 μm [5,6]. Diversity of bioaerosol communities reaches thousands bacterial and fungal genera with tens or hundreds dominant genera, usually including pathogenic microorganisms [7]. Exposure to these airborne bio-particles may pose potential health risks to humans and animals, which include respiratory infections, allergies, and other infectious diseases [8].

Besides the geographical factors, contemporary research indicated that bioaerosols are also influenced by air quality [9], including humidity, ozone (O<sub>3</sub>), sulfur dioxide (SO<sub>2</sub>) and other atmospheric pollutants. Nowadays, O<sub>3</sub> is one of the important air pollutants, exhibits the capability to oxidize all types of organic and inorganic compounds [10]. Thus, O<sub>3</sub> can directly and indirectly interact with bioaerosols due to its ability to capture electrons from other molecules and release them to promote further chemical reactions [11,12]. Direct impacts include the absorption of O<sub>3</sub> by target molecules on bioaerosols, while indirect effects focus on ozonation of water to produce reactive species [13]. At atmospheric O<sub>3</sub> pollution concentrations of tens to hundreds of μg·m<sup>-3</sup>, O<sub>3</sub> has demonstrated remarkable effectiveness in reacting with various types of airborne pathogens, particularly in high humidity conditions (RH > 80%) [14]. Thus, the characteristics of bioaerosols would be significantly influenced by O<sub>3</sub>.

Recently, associations of air pollutions with their health risks have been intensively investigated [5,15]. However, the specific studies on relationships of reactive O<sub>3</sub> with pathogenic bioaerosols have not comprehensively summarized. In this critical mini review, we have highlighted recent research progresses on the mechanisms of atmospheric O<sub>3</sub> mediated characteristics change of bioaerosols, including the influences of O<sub>3</sub> on



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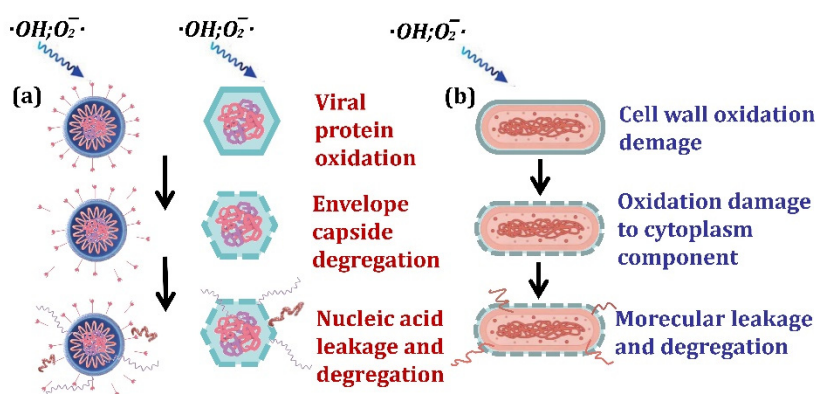
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biological components, bio-activity, size distributions and community structures (concentrations) of bioaerosols, as well as their effects on human health risk in modern scientific cognition, which may alert the public to properly control O<sub>3</sub> and bioaerosol pollutions simultaneously.

## 2. High Concentration Atmospheric O<sub>3</sub> Affects the Size Distribution and Community Structure of Bioaerosols

The World Health Organization (WHO) has established a guideline value of 100 µg·m<sup>-3</sup> as the maximum 8 h average concentration for O<sub>3</sub> (The Grade II standards of the Chinese NAAQS is 160 µg·m<sup>-3</sup> for O<sub>3</sub>), which can induce the mucosal irritation of the human nose and throat [16]. At high concentration of O<sub>3</sub> pollution episodes with >100 µg·m<sup>-3</sup>, O<sub>3</sub> can destroy bioaerosols leading to microorganisms' inactivation, as it is a powerful oxidant [17].

O<sub>3</sub> exhibits the capability to oxidize essential enzymes involved in glucose decomposition and also directly react with and dismantle the organelles, DNA, and RNA of bacteria and fungi, disrupting their metabolic processes and resulting in their demise [18]. O<sub>3</sub> may also exert an inhibitory effect on microorganisms due to its strong oxidizing effect, like oxidizing bacterial cell wall and cytoplasmic membrane, leading to cell lysis and death through damaging the membrane components such as glycoproteins, glycolipids, or amino acids of the nucleic acids [19]. Moreover, O<sub>3</sub> possesses the ability to penetrate cell membranes and target lipoproteins in the outer membrane and lipopolysaccharides in the inner membrane, causing bacterial cell permeability to distort, dissolve, and ultimately perish [16]. On the other hand, O<sub>3</sub> also reacts with other air pollutants including organisms, volatile organic compounds (VOCs), NO<sub>x</sub>, particulate matter (PM) and H<sub>2</sub>O, generating reactive oxygen species (ROSs) like •OH, •O<sub>2</sub><sup>-</sup>, •OOR and so on [19]. These ROSs are also able to inactivate bioaerosols by damaging their proteins or cell wall, resulting in nucleic acid leakage and further degradation [20,21]. As shown in Figure 1, these resulting ROSs can strongly destroy various bacterial cellular tissues and virus [14,22], resulting in reducing bio-activity or death of microorganisms on bioaerosols.



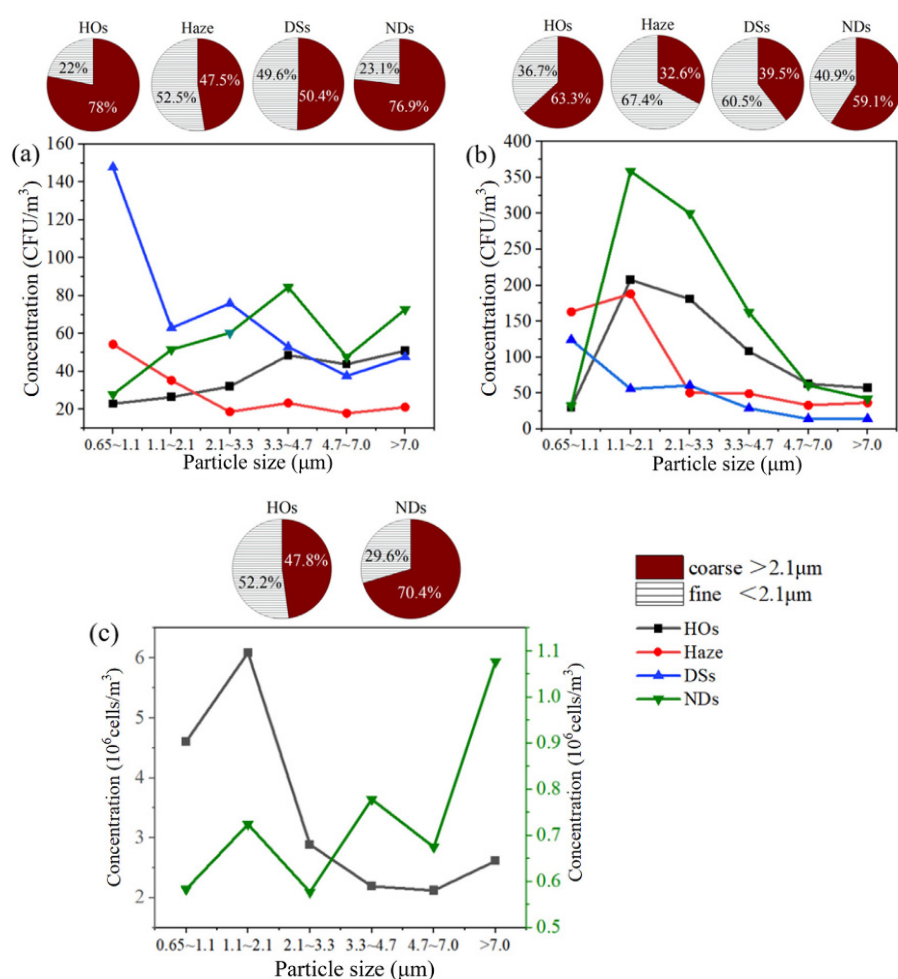
**Figure 1.** Inactivation mechanisms of virus (a) and bacterium (b) with ozone. Reactive oxygen species (ROSs) such as hydroxyl radicals (•OH) and superoxide anion-radicals (•O<sub>2</sub><sup>-</sup>) are produced via O<sub>3</sub> interactions with H<sub>2</sub>O. Reproduced with permission. Copyright 2023, Springer Nature.

Due to the bactericidal property of O<sub>3</sub>, it is easy to consider that exposing to high concentration of O<sub>3</sub> (>100 µg·m<sup>-3</sup>) may lead to reducing concentration of bioaerosols [23]. Actually, as different types of microorganisms on bioaerosols show various tolerance to O<sub>3</sub> (some still alive, while others are inactivated), leading to evolution of community structure, microorganism diversity and altering of size distribution of bioaerosols when exposed to high concentration of O<sub>3</sub>. For fungal aerosols, at high concentration of O<sub>3</sub> pollution, concentration of fungal aerosol decreased from ~950 to ~650 CFU·m<sup>-3</sup> at normal days (14.2 ± 13.7 µg·m<sup>-3</sup>) and O<sub>3</sub> pollution days (102.3 ± 66.2 µg·m<sup>-3</sup>), respectively, and for bacterial aerosol decreased from ~350 to ~210 CFU·m<sup>-3</sup> at normal days and O<sub>3</sub> pollution days, respectively [24]. The bacteria attached to dust on bioaerosols are mainly gram-positive bacteria, which have a stronger tolerance to high O<sub>3</sub> concentration environments than gram-negative bacteria [24]. What should be noted that antibiotic-resistant bacteria (ARB) on bioaerosols are found to be more resistant to O<sub>3</sub> than antibiotic-susceptible bacteria (ASB), indicating that O<sub>3</sub> may increase the proportions of ARB on bioaerosols. Berrington and Pedler found that in contrast to methicillin-sensitive *Staphylococcus aureus* (MSSA), methicillin-resistant *Staphylococcus aureus* (MRSA) strain showed better resistance to high O<sub>3</sub> concentration (100–120 µg·m<sup>-3</sup>) [14]. The number of average survivors of MSSA and MRSA after exposure to 100–120 µg·m<sup>-3</sup> O<sub>3</sub> for 4 h was 3.9 and

31.7, respectively, showing methicillin-resistant MRSA shows better tolerance to  $O_3$ . The results indicate that ARBs on bioaerosols remain alive when exposed to high concentration of  $O_3$ .

Even for virus aerosols, Alimohammadi and Naderi stated that enveloped viruses such as SARS-CoV-2 are less persistent against  $O_3$  molecules (disinfection concentration of  $O_3$   $500 \mu\text{g}\cdot\text{m}^{-3}$ ), compared to non-enveloped viruses [14]. Zuo et al. used redundancy analysis (RDA) and partial RDA (pRDA) to assess the contribution of environmental factors (air pollutants and meteorological conditions) to variation of airborne microorganisms. They found that air pollutants including  $O_3$  played more important role in bacterial community and variations of airborne microorganisms than meteorological conditions [25,26].

After persistent and severe oxidations by high concentration of  $O_3$ , the sensitive bioaerosols eventually die, leading to change of size distribution/diversity and variation of community structure of themselves in some areas [25]. Yang et al. [24] found that high concentration of  $O_3$  ( $102.3 \pm 66.2 \mu\text{g}\cdot\text{m}^{-3}$ ) showed a trend of gradual decrease coarse to fine particle size of bioaerosols, with peak values of  $3.3\text{--}4.7 \mu\text{m}$ . It can be seen from Figure 2, fine culturable bacteria and fungal aerosols ( $<2.1 \mu\text{m}$ ) in  $O_3$  polluted days are similar with normal days at around 20% and 40%, respectively. (HOs is high concentration of  $O_3$  days  $>100 \mu\text{g}\cdot\text{m}^{-3}$ , NDs is normal days). However, fine aerosols of total airborne microbe in HOs were  $\sim 52.2\%$  among all microorganisms, which was much higher than that in NDs ( $\sim 29.6\%$ ), indicating that 20% increase in fine particles and most microorganisms can exist in fine particles under high concentration of  $O_3$  [24]. Similarly, Hao et al. [27] also found that a relatively larger decrease in viability over time for bio-particles within the larger size of  $0.5\text{--}1.0 \mu\text{m}$  under ozone and UV exposure conditions, while no such trend obtained for the smallest bio-particle size range of  $0.3\text{--}0.5 \mu\text{m}$  on bioaerosols. These results give certain evidences that  $O_3$  may promote the reduction of size distribution of bioaerosols, as the viability was higher for smaller particle sizes than for the larger ones.



**Figure 2.** The size distribution of bioaerosols under different types of pollution (HO is high concentration of  $O_3$  days  $>100 \mu\text{g}\cdot\text{m}^{-3}$ , NDs is normal days): (a) airborne bacteria; (b) airborne fungi; (c) total airborne microbe. Reproduced with permission. Copyright 2024, IOP Publishing [24].

As compared with in normal days (concentration of  $O_3 < 20 \mu\text{g}\cdot\text{m}^{-3}$ ), fungi, bacteria and virus on bioaerosols exposure to high concentration ( $>100 \mu\text{g}\cdot\text{m}^{-3}$ ) of  $O_3$  in HOs days, so that the community structure (species  $O_3$  tolerance increase), concentration (decrease), and size distribution (fine bioaerosols increase) of bioaerosols are really modified due to strong oxidization of  $O_3$ . These investigations alert the public that bioaerosols oxidized by high concentration of  $O_3$  may significantly change the concentrations, sizes, categories and community structures of bioaerosols.

### 3. Low Concentration Atmospheric $O_3$ Affects the Biologic Components and Activity of Bioaerosols

At low concentration of atmospheric  $O_3$  condition ( $20\text{--}100 \mu\text{g}\cdot\text{m}^{-3}$ ),  $O_3$  can also target to membrane glycoproteins, glycolipids, and amino acids through modifying the basic chemical structures in nucleic acids and influencing the sulfhydryl groups of vital enzymes, disrupting normal cellular activity. Although the morphology changes of airborne microorganisms caused by  $O_3$  was not so much obvious, there were some researchers claimed that the cell membrane of microbes could be damaged when exposed to high RH and low concentration of  $O_3$  [22].

Actually, the relationship between concentration of microorganisms on bioaerosols and low concentration of  $O_3$  ( $20\text{--}100 \mu\text{g}\cdot\text{m}^{-3}$ ) is still under debate. Some researchers reported that  $O_3$  ( $79.7\text{--}87.6 \mu\text{g}\cdot\text{m}^{-3}$ ) was negatively correlated with many microorganisms, possibly because the toxicity of  $O_3$  inhibits the growth of microorganisms in atmosphere [28], while two works claimed that the total number of Operational taxonomic units (OTUs) was positively correlated with atmospheric  $O_3$  concentration ( $10\text{--}90 \mu\text{g}\cdot\text{m}^{-3}$ ) [29,30].

For bacterial aerosols, at low concentration of  $O_3$  ranging from  $10\text{--}90 \mu\text{g}\cdot\text{m}^{-3}$ , the total number of bacterial OTUs was positively correlated with  $O_3$  concentration ( $R^2 = 0.15$ ) (from 5200 to 5600 CFU $\cdot\text{m}^{-3}$ , 8% increase), indicating that  $O_3$  may promote bacteria growth to some extent, while the relative abundance of pathogenic bacteria was negatively correlated with  $O_3$  ( $R^2 = 0.10$ ) concentration (decreasing from 6% to 4%) [29].

For fungi aerosols, some studies stated that negative relationship was obtained between  $O_3$  and fungal levels [31], which indicating inactivation of fungi on bioaerosols by  $O_3$  even at low concentration of  $O_3$  ( $<100 \mu\text{g}\cdot\text{m}^{-3}$ ). At very low concentration of  $O_3$  ( $5\text{--}6 \mu\text{g}\cdot\text{m}^{-3}$ ), there are some studies found that microorganisms belonging to *phylum Ascomycota* are not especially sensitive to  $O_3$  [32], illustrating that the concentration of specific fungi on bioaerosols didn't being inactivated with increase of  $O_3$ . Actually, different types of fungi show variant tolerance to low concentration of  $O_3$ . Recently, some studies found that specific species were negatively correlated with *Penicillium* but positively correlated with *Alternaria* [32] for harmful organisms being oxidized by  $O_3$ , leaving bioaerosols' growth.

As weak interaction of low concentration  $O_3$  with microorganisms, a slight variation of chemical compositions of microorganisms on bioaerosols is not easily detected. Recently, the scientists have developed Raman spectroscopy strategy for detecting surface chemical composition of a single microorganism before and after  $O_3$  exposure [33]. They investigated Raman spectra of seven different single-trapped fungal aerosol (like *Penicillium camembertii*) in lab, and found that specified several individual chemical function groups of bioaerosols such as lipids and proteins that undergo chemical reactions with  $O_3$ . By comparing Raman spectra of bioaerosols, some new specific characteristic peaks and the change of the ratio of lipid–protein in seven fungus materials can be found after interaction with  $O_3$  for 40 min, indicating that the change of biological compositions of microorganisms on bioaerosols. Similarly, Pan et al. [34,35] reported that both the absorption and emission spectra of aromatic amino acids (tryptophan, tyrosine, and phenylalanine) of bioaerosols were changed by  $O_3$  during oxidation process. Tryptophan, tyrosine, and phenylalanine are main UV fluorescent molecules in bioaerosols, and their fluorescence dramatically decreased at UV but reversely increased at visible range. These results indicate that the biological components (lipids, proteins, DNA and amino acids) of microorganisms on bioaerosols are changed after exposure to  $O_3$ , leading to altering biological activity (metabolism, respiration, mobile performance, reproductive vitality) of microorganisms on bioaerosols.

However, even at low concentration of  $O_3$  pollution, with high relative humidity (RH), the oxidizability of  $O_3$  can be promoted notably. Griffiths et al. claimed that no viability decrease of *P. expansum* spores was found with low concentration of  $O_3$  at low RH, while significant decline can be seen with low concentration of  $O_3$  at high RH, indicating that the reaction of moisture and  $O_3$  seems to enhance the inactivation effects of microorganisms on bioaerosols. For examples, MS2 is found to be more sensitive to  $O_3$  than BtAH ( $10 \mu\text{g}\cdot\text{m}^{-3}$  with RH of 81%) [36]. The reasons are the reactions of  $O_3$  with  $H_2O$  and some low molecular weight hydrocarbons producing large amounts of ROSs and intermediates which maybe toxic to microorganisms on bioaerosols.

In one word, although low concentration of  $O_3$  may not completely inactivate bioaerosols as high concentration of  $O_3$ , owing to its weak oxidation and small possibility of contact with microorganisms, some minor changes of bioaerosols were also really observed. Newly appearing groups and modified lipid-protein ratios

can be found from various microorganisms on bioaerosols when exposure to low concentration of O<sub>3</sub>, which may also influence the biological activity of bioaerosols.

#### 4. Human Health Risks of Ozone-Mediated Bioaerosols

As bioaerosols contains pathogenic microorganisms, which may pose human health risks for example allergy, asthma, infectious diseases and so on. The potential health risks caused to humans mainly depend on the pathogenicity of bioaerosols or potential immunity of human beings, which can be affected by community structures, size distributions, biological compositions and biological activity of bioaerosols. However, these characteristics of bioaerosols can be affected by atmospheric O<sub>3</sub> as discussed above, leading to changes of health effects of them to humans.

At high concentration of O<sub>3</sub> ( $>100 \mu\text{g}\cdot\text{m}^{-3}$ ), some researchers believed that health risks of bioaerosols reduced due to decrease of concentration of bioaerosols, while others claimed that the health risks of bioaerosols increase because producing more harmful components from bioaerosols when exposed to high concentration of O<sub>3</sub>. Some specific microorganisms on bioaerosols are easily inactivated by O<sub>3</sub>, resulting in the total concentration of bioaerosols shows even an order of magnitude decrease. As the pathogenicity of the specific microorganisms and the compounds produced on bioaerosols are reduced, inactivation of these microorganisms by high concentration of O<sub>3</sub> may potentially reduce the health risks of bioaerosols. On the other hand, some studies have shown that O<sub>3</sub> at appropriate concentrations (5–10%) may chemically react with other harmful substances presented on PM, thereby reducing the harm to microorganisms on bioaerosols as well [36]. Bioaerosols containing pathogenic microorganisms are not inactivated, which may increase health risk of them to humans. Moreover, the size distribution of bioaerosols can be decreased at high concentration of O<sub>3</sub>, which may increase health risk of bioaerosols because that the finer size of bioaerosols is able to deposit on the deeper position of human respiratory system (The size of bioaerosol also affects the efficiency of being inhaled and deposited within human respiratory system) [32]. It is because majority of fine-particle bioaerosols can easily enter human body through the respiratory system and can cause a range of diseases, resulting in higher health risk. Both views make sense, as after interaction with high concentration of O<sub>3</sub>, the community structure and size distribution of bioaerosols are really changed, leading to different targeted cells or organs that they attacked or deposited. Therefore, the health risk of bioaerosols at high concentration of O<sub>3</sub> should be further investigated.

At low concentration of O<sub>3</sub> ( $<100 \mu\text{g}\cdot\text{m}^{-3}$ ), the biological compositions and bio-activity of bioaerosols are also changed to some extent. The dominant opinion is that health risks of bioaerosols increases when being exposed to low concentration of O<sub>3</sub>, as they believe that the low concentration of O<sub>3</sub> cannot inactivate bioaerosols, while the biological composition changes of bioaerosol possibly produce more allergenic and toxic contents of pathogenic microorganisms to humans. Since pollen and fungal spores are able to interact with O<sub>3</sub> pollutants, researchers found that they linked to the climate-change-driven (O<sub>3</sub> pollution) worsening respiratory health effects. The exposure to several contaminants (including bioaerosols and O<sub>3</sub>) in urban settings is linked to severe episodes of asthma attacks and/or exacerbations [37]. More precisely, Tiedemann and Firsching [38] reported that the pathogenicity of rust fungi on bioaerosols could be increased after exposure to low concentration of O<sub>3</sub> ( $20\text{--}70 \mu\text{g}\cdot\text{m}^{-3}$ ). Similarly, Zoran et al. considered that low concentration of O<sub>3</sub> ( $20\text{--}50 \mu\text{g}\cdot\text{m}^{-3}$ ) acts as a COVID-19 virus incubator, being positively correlated with COVID-19 infections and fatalities [39]. As strong oxidant, low concentration of ozone induces oxidative stress of bioaerosol, which also cause antimicrobial resistances [40]. In addition, it has been verified that ozone causes conjugative transfer and transformation of antibiotic resistance genes, which may lead severe health risks of ozone mediated bioaerosols [41].

Actually, at really atmospheric environments, both bioaerosols and O<sub>3</sub> at low concentration pose health risks to humans simultaneously [42]. It is known that WHO standard value is  $100 \mu\text{g}\cdot\text{m}^{-3}$ , however, the total mortality associated with levels greater than  $70 \mu\text{g}\cdot\text{m}^{-3}$  account for 0.26% of the deaths, and it is worth noting that even low levels of O<sub>3</sub> ( $50 \mu\text{g}\cdot\text{m}^{-3}$ ) may contribute to mortality [43]. High concentrations of O<sub>3</sub> exposure stimulate the human respiratory system, damage lung cells, central nervous, cardiovascular systems and aggravate other chronic lung diseases, therefore posing a great threat to human health [44]. O<sub>3</sub> not only affects the characteristics of bioaerosols, but can also be inhaled by humans, indicating that both bioaerosols and O<sub>3</sub> pose health risks to humans. Therefore, the co-exposure to bioaerosols and O<sub>3</sub> pollution, the synergistic effects of O<sub>3</sub> and bioaerosols may significantly increase health risks of humans, which should be systematically investigated in the future.

#### 5. Summary and Prospects

With increased safety concerns of bioaerosols and air pollutants, health risks of O<sub>3</sub> mediated bioaerosols and O<sub>3</sub>-bioaerosols co-exposure are under investigated. In heavy O<sub>3</sub> pollution ( $>100 \mu\text{g}\cdot\text{m}^{-3}$ ) with strong interaction

between bioaerosols and O<sub>3</sub> molecular, some microorganisms on bioaerosols are inactivated, leading to concentration and size reduction with change of community structure. The total concentration of microorganisms shows 30% decrease and fine particle size of bioaerosols increase 20% among total bioaerosols due to partially being inactivated by O<sub>3</sub>. In addition, community structures and diversity of bioaerosols change due to different types of microorganisms have varied tolerance to O<sub>3</sub>. Health risks of O<sub>3</sub>-mediated bioaerosols are complicated. To a certain extent, pathogenic microorganisms on bioaerosols can be inactivated by O<sub>3</sub>, thus reducing health risks of bioaerosols. However, the size distribution of bioaerosols decreases as well, improving the potential for a deeper deposition onto respiratory system, which may also increase health risk to humans.

At relatively low concentration of O<sub>3</sub> pollution (20–100 µg·m<sup>-3</sup>), debating opinions believe that concentration of bioaerosols may increase slightly (8% increase) due to clearance of other harmful organisms that toxic to bioaerosols, as well as decrease (2% in pathogenic microorganisms) because of different tolerances to oxidization by O<sub>3</sub>. Even though, O<sub>3</sub> may not able to effectively react with bioaerosols due to long distance and limited oxidizability, biological compositions including ratio of proteins, lipids, and amino acids of bioaerosols are changed, as well as bio-activity of bioaerosols are decreased. Although microbial vitality of bioaerosols decreases, researchers believe that health risk of O<sub>3</sub>-mediated bioaerosols may rise due to usually more allergenic and toxic contents of pathogenic microorganisms on bioaerosols are exposed.

From what discussed in this review, we know that the characteristics of bioaerosols can be changed when interacting with different concentrations of O<sub>3</sub>. However, the investigation of interactions between bioaerosols and O<sub>3</sub> is insufficient due to complex compositions of bioaerosols and lack of proper detection strategy. We summarized following research gaps that should be noticed in future study.

- (1) **Ignoring of assessing viable but non-culturable bioaerosols.** Most of existing researches only focused on O<sub>3</sub> affecting specific or culturable microorganisms on bioaerosols, remaining a lot of unknown and/or viable but non-culturable microorganisms.
- (2) **Influences of O<sub>3</sub> on bioaerosols have not been systematically and deeply investigated.** Most underlying mechanisms of interaction between O<sub>3</sub> and bioaerosols including dose-response relationship, species abundance, metabolic toxicity and so on have not been investigated.
- (3) **Other components on bioaerosols like pollen, endotoxin, and so on have not been considered.** Complicated components on bioaerosols can also be affected by O<sub>3</sub>, causing complex health risks to human beings.
- (4) **Lack of reliable health risks assessment of O<sub>3</sub>-mediated bioaerosols.** The only concentration of total number of culturable microorganisms on bioaerosols has been considered, which is not reasonable.

Most importantly, as both bioaerosols and O<sub>3</sub> are air pollutions with relatively low concentration that cause health risk to humans and animals, there is a lack of researches to summarize the general rules for accurately evaluating health risk of co-exposure to O<sub>3</sub> and bioaerosols.

As there are a lot of unclears remaining, more efforts should be put on investigating interactions and health risks of O<sub>3</sub> and bioaerosols. First of all, detection and sampling strategy of living bioaerosols should be improved, thus viable but not culturable microorganisms can be considered on bioaerosols. Metagenomes sequencing and other methods can be used to reveal the change of community structure, abundance and other components of O<sub>3</sub>-mediated bioaerosols. Secondly, more laboratory investigations to simulate interactions between O<sub>3</sub> and bioaerosols even other air pollutants are needed to reveal underlying mechanisms. Finally, for health risks, as antibiotic resistance gene on bioaerosols poses health risk to human beings, what should be considered in future study is that O<sub>3</sub>-mediated bioaerosols may promote the horizontal transform of genes. A methodology for assessing health risks of O<sub>3</sub>-mediated bioaerosols and co-exposure of bioaerosols and O<sub>3</sub> also should be urgently built. We believe comprehensive investigations on the interactions of O<sub>3</sub> with bioaerosols to accurately reveal deep mechanisms through advanced technologies are still on the way. This critical mini review may alert the public to reveal the health risks of exposure to bioaerosols and O<sub>3</sub>, as well as properly control O<sub>3</sub> and bioaerosol pollutions.

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## References

1. Amin, H.; Šantl-Temkiv, T.; Cramer, C.; Finster, K.; Real, F.G.; Gislason, T.; Holm, M.; Janson, C.; Jøgi, N.O.; Jogi, R.; et al. Indoor airborne microbiome and endotoxin: Meteorological events and occupant characteristics are important determinants. *Environ. Sci. Technol.* **2023**, *57*, 11750–11766.
2. Li, P.; Li, L.; Yang, K.; Zheng, T.; Liu, J.; Wang, Y. Characteristics of microbial aerosol particles dispersed downwind from rural sanitation facilities: Size distribution, source tracking and exposure risk. *Environ. Res.* **2021**, *195*, 110798.
3. Chen, Y.; Liang, Z.; Li, G.; An, T. Indoor/Outdoor airborne microbiome characteristics in residential areas across four seasons and its indoor purification. *Environ. Int.* **2024**, *190*, 108857.
4. Su, K.; Liang, Z.; Zhang, S.; Liao, W.; Gu, J.; Guo, Y.; Li, G.; An, T. The abundance and pathogenicity of microbes in automobile air conditioning filters across the typical cities of China and Europe. *J. Hazard. Mater.* **2024**, *472*, 134459.
5. Zhang, S.; Liang, Z.; Wang, X.; Ye, Z.; Li, G.; An, T. Bioaerosols in an industrial park and the adjacent houses: Dispersal between indoor/outdoor, the impact of air purifier, and health risk reduction. *Environ. Int.* **2023**, *172*, 107778.
6. Liang, Z.; Yu, Y.; Wang, X.; Liao, W.; Li, G.; An, T. The exposure risks associated with pathogens and antibiotic resistance genes in bioaerosol from municipal landfill and surrounding area. *J. Environ. Sci.-China* **2023**, *129*, 90–103.
7. Yue, S.; Li, L.; Xu, W.; Zhao, J.; Ren, H.; Ji, D.; Li, P.; Zhang, Q.; Wei, L.; Xie, Q.; et al. Biological and nonbiological sources of fluorescent aerosol particles in the urban atmosphere. *Environ. Sci. Tech.* **2022**, *56*, 7588–7597.
8. Geng, X.; Nie, C.; Chen, H.; Tang, X.; Wei, M.; Wang, Y.; Gao, H.; Li, D.; Fang, M.; Ju, R.; et al. Nycterohemeral airborne fungal and bacterial communities and health risks of potential pathogens in Shanghai. *Environ. Sci. Atmos.* **2024**, *4*, 190–201.
9. Cariñanos, P.; Foyo-Moreno, I.; Alados, I.; Guerrero-Rascado, J.L.; Ruiz-Peñuela, S.; Titos, G.; Cazorla, A.; Alados-Arboledas, L.; de la Guardia, C.D. Bioaerosols in urban environments: Trends and interactions with pollutants and meteorological variables based on quasi-climatological series. *J. Environ. Manag.* **2021**, *282*, 111963.
10. Wang, H.; Wang, H.; Lu, X.; Lu, K.; Zhang, L.; Tham, Y.J.; Shi, Z.; Aikin, K.; Fan, S.; Brown, S.S.; et al. Increased night-time oxidation over China despite widespread decrease across the globe. *Nat. Geosci.* **2022**, *16*, 217–223.
11. Wang, Y.; Zhao, Y.; Liu, Y.; Jiang, Y.; Zheng, B.; Xing, J.; Liu, Y.; Wang, S.; Nielsen, C.P. Sustained emission reductions have restrained the ozone pollution over China. *Nat. Geosci.* **2023**, *16*, 967–974.
12. Wang, H.; Peng, L.; Li, G.; Liu, H.; Liang, Z.; Zhao, H.; An, T. Enhanced catalytic ozonation inactivation of bioaerosols by MnO<sub>2</sub>/Ni foam with abundant oxygen vacancies and O<sub>3</sub> at atmospheric concentration. *Appl. Catal. B Environ.* **2024**, *344*, 123675.
13. Kakaei, K.; Padervand, M.; Akinay, Y.; Dawi, E.; Ashames, A.; Saleem, L.; Wang, C. A critical mini-review on challenge of gaseous O<sub>3</sub> toward removal of viral bioaerosols from indoor air based on collision theory. *Environ. Sci. Pollut. Res.* **2023**, *30*, 84918–84932.
14. Truyols-Vives, J.; Botella-Grau, S.; Mercader-Barceló, J.; Baldoví, H.G. Antimicrobial activity of safe concentrations of ozone, hydrogen peroxide, and triethylene glycol in air and surfaces. *Environ. Sci. Atmos.* **2024**, *4*, 620–633.
15. Liang, Z.; Yu, Y.; Ye, Z.; Li, G.; Wang, W.; An, T. Pollution profiles of antibiotic resistance genes associated with airborne opportunistic pathogens from typical area, Pearl River Estuary and their exposure risk to human. *Environ. Inter.* **2020**, *143*, 105934.
16. Liu, Z.; Xiao, X.; Jiang, C.; Wang, Y.; He, J. Assessment of the air disinfection effect of low-concentration ozone in a closed environment. *Build. Environ.* **2023**, *244*, 110747.
17. Yao, M.; Zhang, L.; Ma, J.; Zhou, L. On airborne transmission and control of SARS-CoV-2. *Sci. Total. Environ.* **2020**, *731*, 139178.
18. Premjit, Y.; Sruthi, N.U.; Pandiselvam, R.; Kothakota, A. Aqueous ozone: Chemistry, physiochemical properties, microbial inactivation, factors influencing antimicrobial effectiveness, and application in food. *Compr. Rev. Food Sci. Food Saf.* **2022**, *21*, 1054–1085.
19. Afonso, N.F.; Pires, J.C. Characterization of surface ozone behavior at different regimes. *Appl. Sci.* **2017**, *7*, 944.
20. Li, G.; Liu, X.; An, T.; Yang, H.; Zhang, S.; Zhao, H. Photocatalytic and photoelectrocatalytic degradation of small biological compounds at TiO<sub>2</sub> photoanode: A case study of nucleotide bases. *Catal. Today* **2015**, *242*, 363–371.
21. Sun, H.; Li, G.; Nie, X.; Shi, H.; Wong, P.K.; Zhao, H.; An, T. Systematic approach to in-depth understanding of photoelectrocatalytic bacterial inactivation mechanisms by tracking the decomposed building blocks. *Environ. Sci. Technol.* **2014**, *48*, 9412–9419.
22. Wang, H.; Peng, L.; Li, G.; Zhang, W.; Liang, Z.; Zhao, H.; An, T. Photocatalytic ozonation inactivation of bioaerosols by NiFeOOH nanosheets in situ grown on nickel foam. *Appl. Catal. B Environ.* **2023**, *324*, 122273.
23. Pironti, C.; Moccia, G.; Motta, O.; Boccia, G.; Franci, G.; Santoro, E.; Capunzo, M.; De Caro, F. The influence of microclimate conditions on ozone disinfection efficacy in working places. *Environ. Sci. Poll. Res.* **2021**, *28*, 64687–64692.
24. Yang, Y.; Yang, L.; Hu, X.; Shen, Z. Characteristics of bioaerosols under high-ozone periods, haze episodes, dust storms,



- and normal days in Xi'an, China. *Particuology* **2024**, 9, 140–148.
25. Gong, J.; Qi, J.; Beibei, E.; Yin, Y.; Gao, D. Concentration, viability and size distribution of bacteria in atmospheric bioaerosols under different types of pollution. *Environ. Pollut.* **2020**, 257, 113485.
26. Zuo, Z.; Pan, Y.; Huang, X.; Yuan, T.; Liu, C.; Cai, X.; Xu, Z. Seasonal distribution of human-to-human pathogens in airborne PM<sub>2.5</sub> and their potential high-risk ARGs. *Front. Microbiol.* **2024**, 15, 1422637.
27. Hao, W.; Huang, Y.W.; Wang, Y. Bioaerosol size as a potential determinant of airborne *E. coli* viability under ultraviolet germicidal irradiation and ozone disinfection. *Nanotechnology* **2024**, 35, 145702.
28. Li, Z.; Lu, J.; Tong, Y.; Li, S.; He, F. Differences in microbial community composition and factors affecting different particulate matter during autumn in three cities of Xinjiang, China. *Sci. Total. Environ.* **2023**, 866, 161275.
29. Liu, H.; Zhang, X.; Zhang, H.; Yao, X.; Zhou, M.; Wang, J.; He, Z.; Zhang, H.; Lou, L.; Mao, W.; et al. Effect of air pollution on the total bacteria and pathogenic bacteria in different sizes of particulate matter. *Environ. Pollut.* **2018**, 233, 483–493.
30. Góralska, K.; Lis, S.; Gawor, W.; Karuga, F.; Romaszko, K.; Brzezińska-Lasota, E. Culturable filamentous fungi in the air of recreational areas and their relationship with bacteria and air pollutants during winter. *Atmosphere* **2022**, 13, 207.
31. Cordero, J.M.; Núñez, A.; García, A.M.; Borge, R. Assessment and statistical modelling of airborne microorganisms in Madrid. *Environ. Pollut.* **2021**, 269, 116124.
32. Wang, Y.; Qi, J.; Han, C.; Zhang, T.; Zhang, D. Microbial characteristics of culturable fungi and bacteria in aerosol particles of a coastal region. *Aerobiologia* **2020**, 36, 507–525.
33. Ai, Y.; Wang, C.; Pan, Y.L.; Videen, G. Characterization of single fungal aerosol particles in a reactive atmospheric environment using time-resolved optical trapping-raman spectroscopy (OT-RS). *Environ. Sci. Atmos.* **2022**, 2, 591–600.
34. Pan, Y.L.; Santarpia, J.L.; Ratnesar-Shumate, S.; Corson, E.; Eshbaugh, J.; Hill, S.C.; Chatt, Williamson, C.; Coleman, M.; Bare, C.; Kinahan, S. Effects of ozone and relative humidity on fluorescence spectra of octapeptide bioaerosol particles. *J. Quant. Spectrosc. Radiat. Transf.* **2014**, 133, 538–550.
35. Pan, Y.L.; Kalume, A.; Wang, C.; Santarpia, J. Atmospheric aging processes of bioaerosols under laboratory-controlled conditions: A review. *J. Aerosol. Sci.* **2021**, 155, 105767.
36. Ratnesar-Shumate, S.; Pan, Y.L.; Hill, S.C.; Kinahan, S.; Corson, E.; Eshbaugh, J.; Santarpia, J.L. Fluorescence spectra and biological activity of aerosolized bacillus spores and MS2 bacteriophage exposed to ozone at different relative humidities in a rotating drum. *J. Quant. Spectrosc. Radiat. Transf.* **2015**, 153, 13–28.
37. D'Amato, G.; Annesi-Maesano, I.; Biagioni, B.; Lancia, A.; Cecchi, L.; D'Ovidio, M.C.; D'Amato, M. New developments in climate change, air pollution, pollen allergy, and interaction with SARS-CoV-2. *Atmosphere* **2023**, 14, 848.
38. Tiedemann, A.V.; Firsching, K.H. Interactive effect of elevated ozone and carbon dioxide on growth and yield of leaf rust infected versus non infected wheat. *Environ. Pollut.* **2000**, 108, 357–363.
39. Zoran, M.A.; Savastru, R.S.; Savastru, D.M.; Tautan, M.N. Assessing the relationship between ground levels of ozone (O<sub>3</sub>) and nitrogen dioxide (NO<sub>2</sub>) with coronavirus (COVID-19) in Milan, Italy. *Sci. Total. Environ.* **2020**, 740, 140005.
40. Bai, C.; Cai, Y.; Sun, T.; Li, G.; Wang, W.; Wong, P.K.; An, T. Mechanism of antibiotic resistance spread during sub-lethal ozonation of antibiotic-resistant bacteria with different resistance targets. *Water Res.* **2024**, 259, 121837.
41. Alexander, J.; Knopp, G.; Dötsch, A.; Wieland, A.; Schwartz, T. Ozone treatment of conditioned wastewater selects antibiotic resistance genes, opportunistic bacteria, and induce strong population shifts. *Sci. Total Environ.* **2016**, 559, 103–112.
42. Wang, Y.; Yang, Y.; Yuan, Q.; Li, T.; Zhou, Y.; Zong, L.; Wang, M.; Xie, Z.; Ho, H.C.; Gao, M.; et al. Substantially underestimated global health risks of current ozone pollution. *Nat. Commun.* **2025**, 16, 102.
43. Zong, L.; Yang, Y.; Xia, H.; Gao, M.; Sun, Z.; Zheng, Z.; Li, X.; Ning, G.; Li, Y.; Lolli, S. Joint occurrence of heatwaves and ozone pollution and increased health risks in Beijing, China: Role of synoptic weather pattern and urbanization. *Atmos. Chem. Phys.* **2022**, 22, 6523–6538.
44. Carvalho, R.B.; Marmett, B.; Dorneles, G.P.; da Silva, I.M.; Romão, P.R.T.; da Silva Júnior, F.M.R.; Rhoden, C.R. O<sub>3</sub> concentration and duration of exposure are factors influencing the environmental health risk of exercising in Rio Grande, Brazil. *Environ. Geochem. Health* **2022**, 44, 2733–2742.